

# DENOThe Meeting 2019

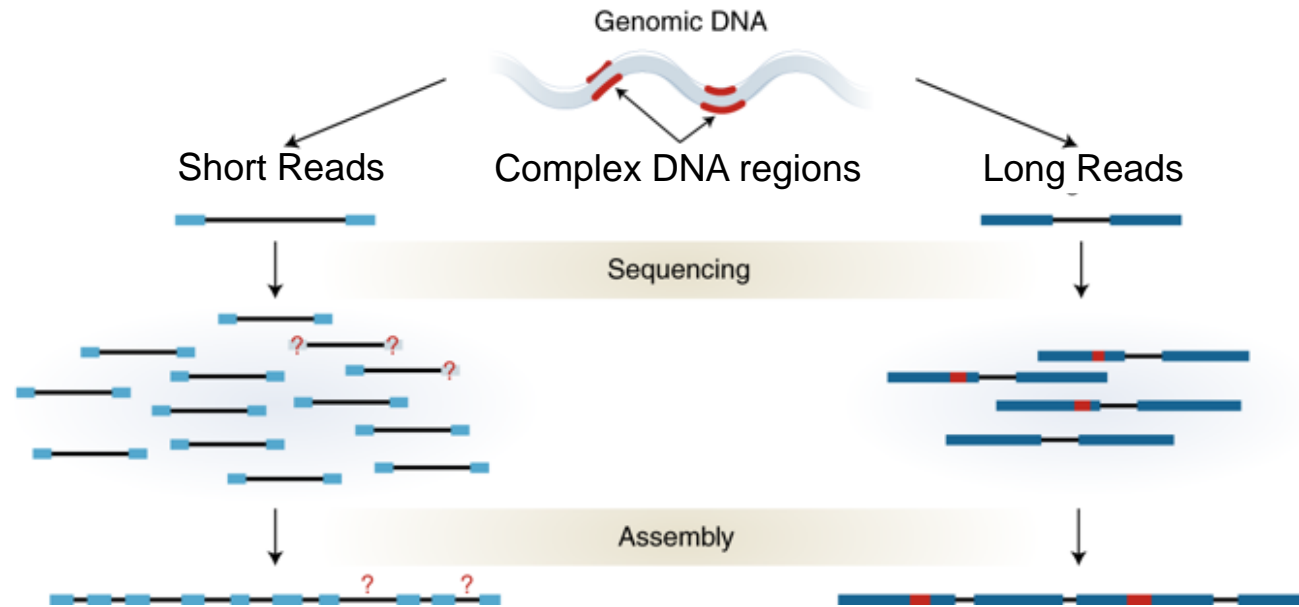
## Sequenziamento Nanopore applicazioni in onco- ematologia

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# Third Generation Sequencing

- The majority of cancers are driven by genomic alterations defined as structural variants (SVs), including translocations, inversions and copy number variations (CNVs).
- SVs detection tools rely on the so-called Second-Generation Sequencing (SGS) platforms, these technologies face inaccuracy and limitations in detecting some classes of SVs.
- In this scenario, the advent of new long reads (up to 20 Kb) -based Third-Generation Sequencing (TGS) technologies might allow to explore those genomic regions uncovered by SGS.





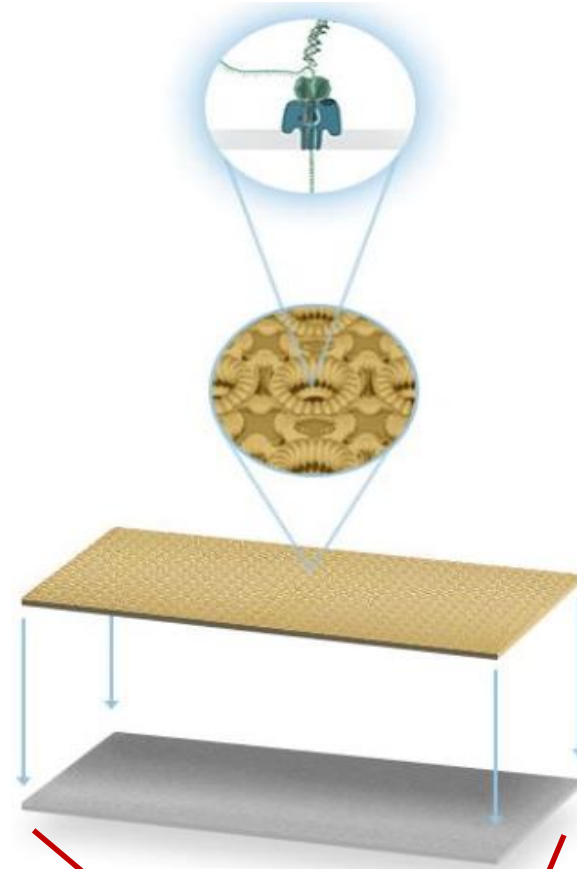
# Nanopore Sequencing – How it works?

## Nanopore

A sampling ionic current is applied through each pore by setting a voltage across this membrane, the translocation of a strand of DNA through the nanopore creates a characteristic disruption in current as the oligonucleotides pass through the pore in different combinations.

## Array Chip

Each microscaffold on the sensor array chip contains an individual electrode, allowing for multiple nanopore experiments to be performed in parallel.

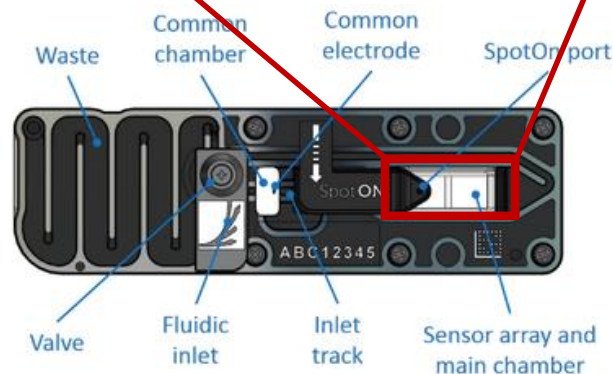


## Array of Microscaffolds

An array of microscaffolds holds the membrane in which the nanopore are embedded. This keep the membrane stable during shipping and usage.

## Application Specific Integrated Circuit (ASIC)

Each nanopore is controlled and measured by an individual channel on a corresponding, bespoke ASIC.



## Flow-cell 9.4.1 RevD

The Flow Cells contain the proprietary sensor array, Application-Specific Integrated Circuit (ASIC) and nanopores that are needed to perform a complete single-molecule sensing experiment.

# Nanopore Platforms



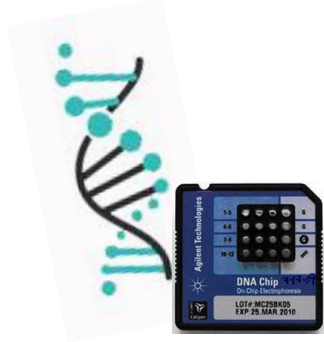
- **De novo AML**
- **Secondary AML**
- **MPN**
- **Prenatal Diagnosis**

Overview	MinION	GridION
Yield per flow cell, DNA, cDNA	50 Gb	50 Gb
Number of Flow cell per device	1	5
Yield per device	50 Gb	250 Gb
Application	Whole Genome/Exomes Targeted Sequencing Whole Transcriptomes (cDNA) Smaller transcriptomes (direct RNA)	Larger Genome Whole Transcriptomes (cDNA or direct RNA)

# Nanopore Sequencing – Workflow



Samples collection



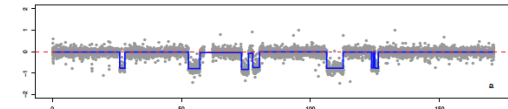
gDNA extraction



Library prep.



GridION sequencing



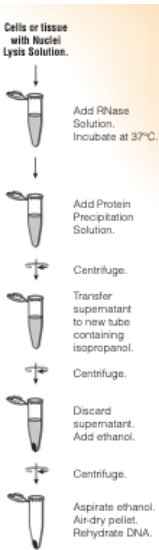
Nano-GLADIATOR analysis

Raw Reads

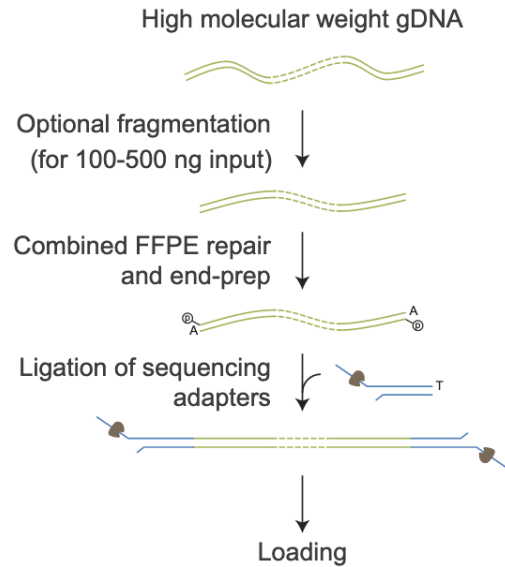
Alignment to Reference Genome

CNVs calling

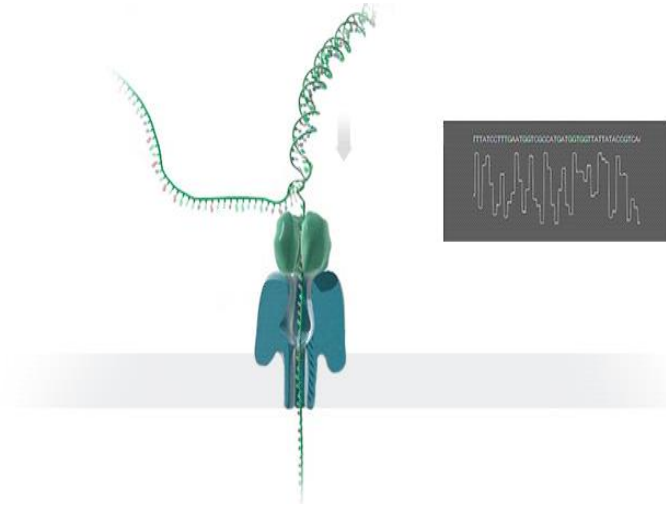
2 h



90 min



60 min



48 h

- **De novo AML**
- **Secondary AML**
- **TN MPN**
- **Prenatal Diagnosis**

# Nanopore Sequencing – Advantages and Limitation



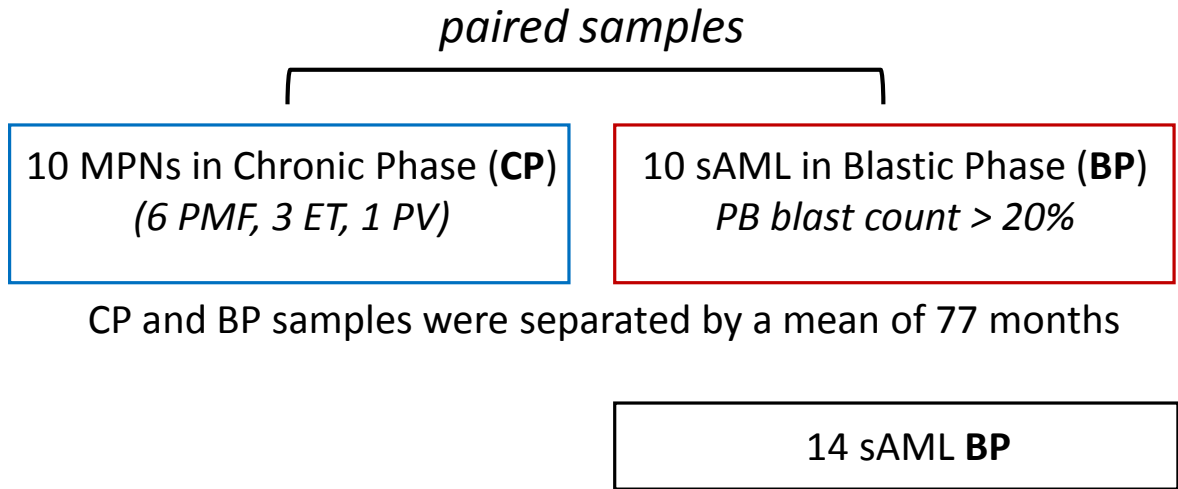
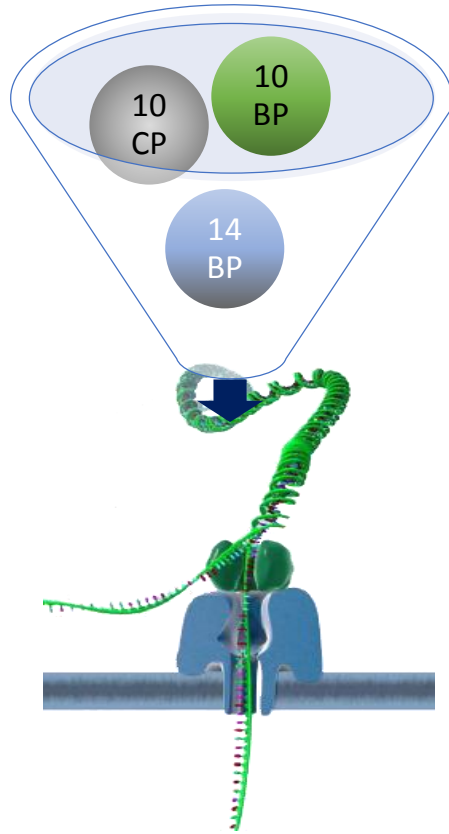
- Low cost
- Sequencing of native DNA molecules – no bias and loss of valuable information
- Sequencing of Repetitive DNA regions
- Time



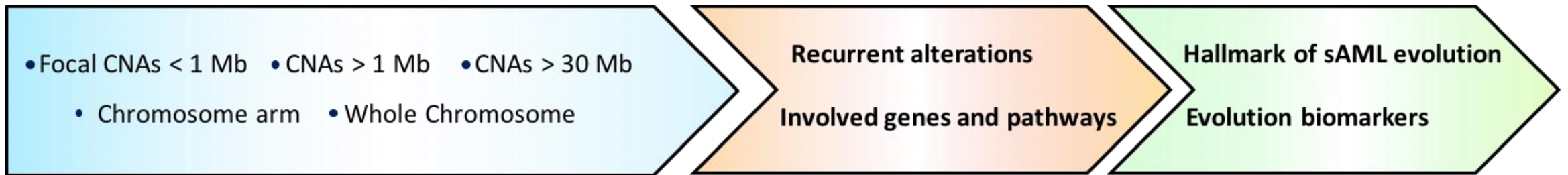
- Error Rate ~ 5%
- DNA quantity ~ 700 ng



# Nanopore Sequencing – Application on sAML

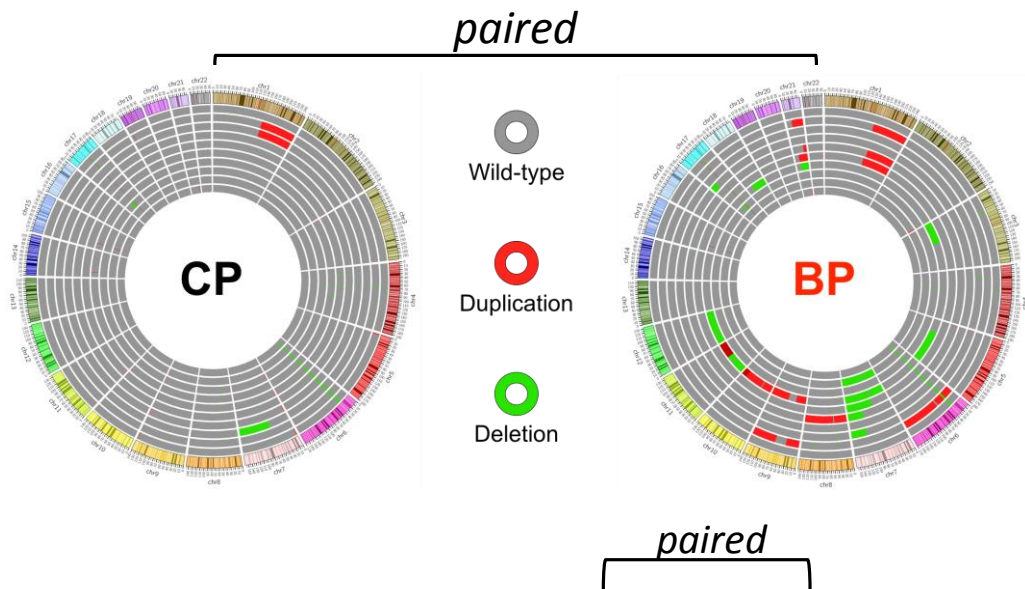


## CNAs identification and categorization <sup>[1,2,3]</sup>

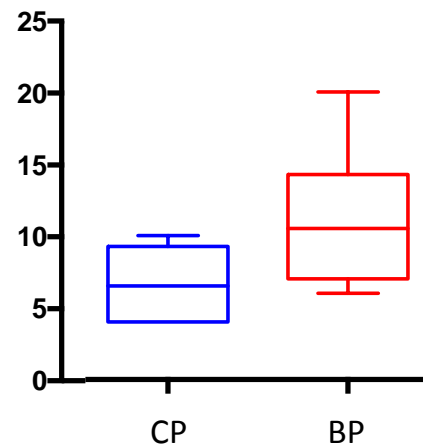




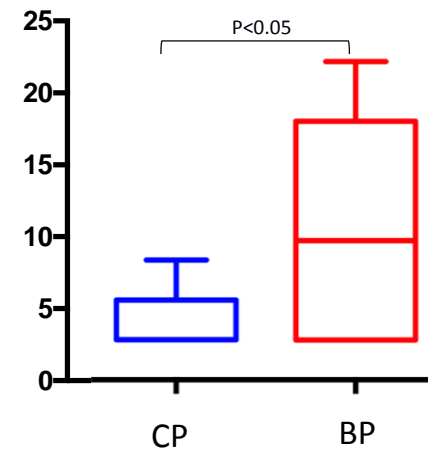
# Nanopore Sequencing– sAML Preliminary Results



Mean CNAs Number

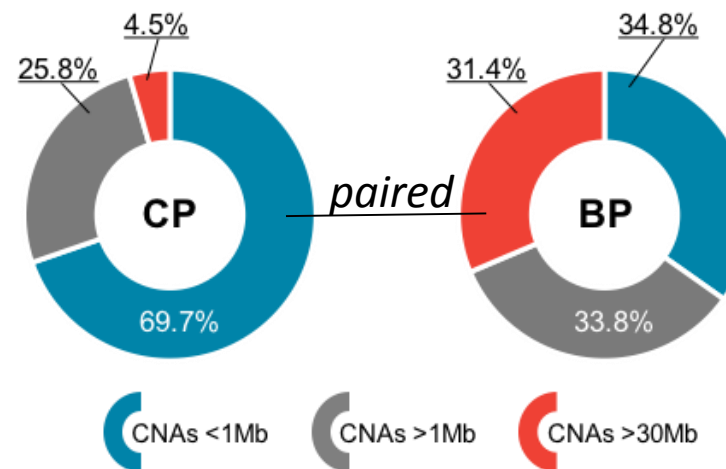


Mean CNAs>30Mb Number



Variable	CP	BP
Total CNAs mean number	6.6±2.6	11.1±4.4
Focal CNAs (%)	69.7	34.8
>1Mb CNAs (%)	25.8	33.8
>30 Mb CNAs (%)	4.5	31.4
Samples with $\geq 1$ arm altered	20% (2 of 10)	40% (4 of 10)
Samples with $\geq 1$ chr altered	none	40%
Mean VAF	0.81	0.79
Mean bp involved	7.07x10 <sup>6</sup>	16.64x10 <sup>6</sup>

CNAs length distribution (%)



# Nanopore Sequencing – To Sum Up

